

# Enhanced imaging properties of a Gd<sup>III</sup> complex with unusually large relaxivity

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## Abstract

MRI contrast properties of self-aggregated nanoparticles made from Gd<sup>III</sup> and 1,2,4,5-tetrakis(pyrazol-1-yl-methyl-3-carboxylate)benzene (L) are investigated. Viscosity of GdL<sub>2</sub> suspensions and size-characteristics of GdL<sub>2</sub> nanoparticles allow an estimate of their large rotational correlation time. Moreover, two to three H<sub>2</sub>O molecules are bound, on average, to Gd<sup>III</sup> ions, as deduced from <sup>17</sup>O NMR titration of the Dy<sup>III</sup> analogue. The large relaxivity of the particles, along with the prominent peak in the range 20–60 MHz, are the consequence of these two properties. Longitudinal ( $r_1$ ) and transverse ( $r_2$ ) relaxivities are determined as a function of monomer concentration at 20 °C and 20 MHz. The ratio  $r_1/r_2$  appears to be favorable for MR imaging using  $T_1$ -weighted gradient echo sequences. According to preliminary tests conducted under physiological conditions, the GdL<sub>2</sub> nanoparticles have some potential as contrast agent provided their stability can be increased. © 2003 Elsevier B.V. All rights reserved.

**Keywords:** Gadolinium; Podate; Relaxivity; Magnetic resonance imaging; Hydration number

## 1. Introduction

Trivalent gadolinium is well-known to locally create a strong contrasting effect in magnetic resonance imaging (MRI) by increasing the longitudinal and transverse proton relaxation rates of the water molecules in close contact with it [1]. This technique is finding an increasing number of applications both in medical diagnostics and analyses [2]. Among the various parameters affecting the relaxivity, the most important are [3]: the number of water molecules coordinated onto the paramagnetic ion and their exchange rate; the electronic relaxation rate of the paramagnetic ion, and the rotational correlation time around the Gd<sup>3+</sup>–OH<sub>2</sub> bond. The latter parameter influences significantly the relaxivity profile in the Larmor frequency range 10–100 MHz. Theoretically, the relaxivity versus frequency curve should present a peak of relaxivity, whose theoretical value should culminate at approximately 100–200 mM<sup>−1</sup> s<sup>−1</sup> for large Gd-containing particles [4].

While a first generation of contrast agents has taken advantage of the thermodynamic and kinetic stability of simple chelates such as [Gd(dota)]<sup>−</sup> or [Gd(dtpa)]<sup>2−</sup>, a second

generation of molecules is presently being designed, with the aim of developing smart properties such as specific sensing [5] or targeting [6], or very high relaxivity [7]. In particular, the latter is needed for angiography for which it is necessary to create a strong contrast within a short time, compatible with the retention time of the contrast agent in the blood. The main efforts to design high relaxivity contrast agents have relied on increasing the rotational correlation time by using either specific association of the paramagnetic probe with proteins [8], or its insertion into large edifices such as dendrimers [9], micelles [10], or aggregates with low critical aggregation concentrations. We have chosen the latter approach and have demonstrated that the Gd<sup>III</sup> podates with 1,2,4,5-tetrakis(pyrazol-1-yl-methyl-3-carboxylate)benzene (L) self-aggregate in water to yield nanoparticles with well defined sizes (10, 60 and 280 nm, critical aggregation concentrations:  $cac-1 = 3.5 \times 10^{-5}$  M and  $cac-2 = 10^{-4}$  M) and that suspensions of these particles present a relaxivity which is about ten times larger than that of the first generation contrast agents [11]. In this paper, we determine the average number of water molecules per lanthanide ion, the rotational correlation time  $\tau_R$ , as well as  $r_1$  and  $r_2$  NMRD profiles of {[GdL<sub>2</sub>(H<sub>2</sub>O)<sub>q</sub>]<sup>5−</sup>}<sub>n</sub> suspensions (abbreviated GdL<sub>2</sub>) under MRI experimental conditions, in order to get more insight into their properties and to assess their feasibility for in vivo applications.

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## 2. Experimental

### 2.1. Synthesis and materials

The tetradentate podand L and its complexes were synthesized as previously described [11]. Lanthanide salts were prepared from 99.99% oxide (Rhône-Poulenc) and the corresponding analytical grade acid. Concentrations were determined by complexometric titrations using standardized  $\text{Na}_2\text{H}_2\text{edta}$  solutions in urotropine buffered medium and xylenol orange as indicator.

### 2.2. $^{17}\text{O}$ NMR measurements on the $\text{Dy}^{\text{III}}$ podate

Suspensions of the  $\{\text{DyL}_2(\text{H}_2\text{O})_q\}^{5-}_n$  aggregated podate ( $\text{DyL}_2$ ) were prepared by mixing a solution of L (0.0251 M, pH  $\sim 7$ ) with a freshly prepared solution of  $\text{Dy}(\text{NO}_3)_3$  (0.1525 M) in stoichiometric ratio. The concentrations of the  $\text{DyL}_2$  aqueous suspensions used (1 ml) were in the range  $7 \times 10^{-4}$  to  $5.0 \times 10^{-3}$  M and the pH was adjusted to 7.5.  $\text{Dy}^{\text{III}}$ -induced  $^{17}\text{O}$  chemical shifts ( $\delta$ ) of  $\text{H}_2\text{O}$  from  $[\text{Dy}(\text{H}_2\text{O})_8]^{3+}$  were calibrated versus the metal concentration before determining the shifts of  $\text{DyL}_2$  versus concentration. Measurements were carried out on a DPX-600 Bruker NMR spectrometer ( $B = 14.1$  T, internal reference  $^{17}\text{O}$ -enriched  $\text{CH}_3\text{NO}_2$ ) at both 50 and 25 °C. The average number of  $\text{H}_2\text{O}$  molecules,  $q$ , per  $\text{Dy}^{\text{III}}$  ion was obtained using Eq. (1) [12]:

$$\text{dis} = \frac{q\delta c}{[\text{H}_2\text{O}]_{\text{solv}}} \quad (1)$$

where  $\delta$  is the mean chemical shift of bound water  $^{17}\text{O}$  nucleus (obtained from the calibration curve) and  $c$  the concentration of the complex. Since at the concentrations used  $[\text{H}_2\text{O}]_{\text{solv}}$  is nearly constant, a linear relationship is expected between  $\text{dis}$  and  $c$  with a slope  $q\delta$ .

### 2.3. Viscosity measurements

Viscosity measurements of  $\text{GdL}_2$  suspensions ( $[\text{GdL}_2]_{\text{tot}} = 1.1 \times 10^{-4}$  M) were carried out with an Ostwald capillary viscometer calibrated at 20.0 °C with four fluids (water, ethanol, methanol, heptane). Calibration was performed by using Eq. (2) from flow-rate measurements and knowing the viscosity and density of the four reference fluids.

$$\eta/\rho = At - B/t \quad (2)$$

with  $\eta$  being the viscosity (cps),  $\rho$  the density (g/l),  $t$  the time (s), while  $A$  and  $B$  are adjustable parameters. Viscosities were measured versus  $T$  (280–345 K) in order to estimate the rotation correlation time ( $\tau_R$ ) of the nanoparticles by using the Stokes–Einstein–Debye [13] Eq. (3), giving  $\tau_R$  for spherical particles as a function of the suspension viscosity:

$$\tau_R (\text{s}) = \frac{4\pi r^3 \eta}{3k_B T} \quad (3)$$

with  $r$  (m) being the effective hydrodynamic radius of the particle,  $\eta$  (Pa s) the solution viscosity,  $T$  (K) the temperature, and  $k_B$  ( $\text{Pa m}^3 \text{K}^{-1}$ ) the Boltzmann constant.

### 2.4. Longitudinal ( $r_1$ ) and transverse ( $r_2$ ) proton relaxivity from MRI sequences

Contrast measurements by MRI of  $\text{GdL}_2$  suspensions at pH 7.4 were carried out on a MRI Symphony Siemens device ( $B = 1.5$  T) at 20 °C and 63 MHz proton Larmor frequency. Relaxation times,  $T_i$  ( $i = 1, 2$ ) were obtained by measuring the image contrast (from black to white) versus time after an initial excitation pulse and by adjusting the experimental values with exponential functions  $I_i = I_{0,i}(1 - \exp(-t/T_i))$ , with  $i = 1, 2$ . For  $T_1$  measurements, three time delays were used (350, 1000 and 2000 ms) and relaxivity was readjusted by using the known value for Omniscan®. Only estimates for  $r_1$  could therefore be obtained from such a procedure and were used to confirm the more precise data extracted from NMRD profiles [11]. For  $T_2$  measurements, twelve time delays were used (from 22.5 to 360 ms with 22.5 ms-intervals). Calculated values of  $r_i$  ( $\text{mM}^{-1} \text{s}^{-1}$ ) were obtained by adjusting  $1/T_i$  values versus  $[\text{Gd}]_{\text{tot}}$  (mM) according to:

$$\left(\frac{1}{T_i}\right)_{\text{meas}} = \left(\frac{1}{T_i}\right)_{\text{water}} + r_i [\text{Gd}]_{\text{tot}} \quad (4)$$

## 3. Results and discussion

Following our previous study [11], we have attempted to determine the number of coordinated water molecules per lanthanide ion,  $q$ , in the aggregated nanoparticles using a luminescence lifetime method with  $\text{Ln} = \text{Eu}^{\text{III}}$  and  $\text{Tb}^{\text{III}}$ . Data were however not precise enough so that we could only estimate  $q \approx 3$  for  $\text{TbL}_2$ . In order to get a more accurate value, we have turned to another method based on the  $^{17}\text{O}$  dysprosium-induced NMR shifts ( $\text{dis}$ ). Indeed, these shifts are dominated by the contact contribution (usually  $>85\%$ ) and are almost independent of the nature of the ligand(s) coordinated to  $\text{Dy}^{\text{III}}$ . Therefore, the mean shift per bound water molecule  $\delta$  may be determined from a calibration curve based on the aquo-ion ( $q = 8$ ) and measurement of  $\text{dis}$  versus the complex concentration yields the  $q$  value. Some caution has to be taken since  $q$  may vary with the chelate concentration. In our case, we have found a linear relationship between the  $\text{dis}$  in  $\{\text{DyL}_2(\text{H}_2\text{O})_q\}^{5-}_n$  and the concentration of the aggregated nanoparticles in the range  $7 \times 10^{-4}$  to  $5.0 \times 10^{-3}$  M, meaning that  $q$  remains constant in this concentration range. The number of bound  $\text{H}_2\text{O}$  molecules amounts to  $q = 1.9 \pm 0.3$ . This allows us to get some insight into the coordination environment of the  $\text{Ln}^{\text{III}}$  ion. Assuming a coordination number of 8 for  $\text{Dy}^{\text{III}}$ , the presence of two inner-sphere water molecules implies that the ligand probably acts as a tridentate host. Therefore, since  $\text{Gd}^{\text{III}}$  has the tendency of having a larger

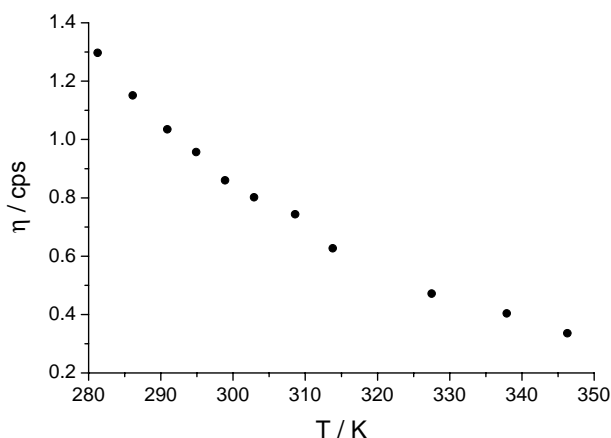


Fig. 1. Viscosity of a GdL<sub>2</sub> suspension versus temperature; [GdL<sub>2</sub>]<sub>tot</sub> =  $1.1 \times 10^{-4}$  M; pH 7.4.

coordination number than Dy<sup>III</sup>, a reasonable estimate for the mean number of coordinated water molecules per Gd<sup>III</sup> ion in the GdL<sub>2</sub> particles lies between 2 and 3.

The nanoparticles have a surface-active effect, inducing a decrease in the solution viscosity as the concentration is increased. The dependence of the solution viscosity upon temperature in the range 281–346 K, at the second critical aggregation concentration [GdL<sub>2</sub>]<sub>tot</sub> =  $1.1 \times 10^{-4}$  M, is presented on Fig. 1. At this concentration, most of the nanoparticles are spherical and have a diameter between 10 and 20 nm, as demonstrated both by light scattering measurements and transmission electron microscopy [11]. Taking this information into account, the rotation correlation time  $\tau_R$  can be estimated, from the viscosity dependence upon temperature, to lie in the range 100 ns–1  $\mu$ s (see Eq. (3)). This is apparently a very large value, about two orders of magnitude larger than those reported, on the basis of NMR measurements, for nano-scale edifices such as dendrimers ( $\tau_R$  = 1–10 ns) or the adduct between a Gd<sup>III</sup> complex with dotp and human serum albumin HSA ( $\tau_R$  = 3 ns, radius  $\sim$ 5 nm) [14]. However, it is known that Eq. (3) provides  $\tau_R$  values one order of magnitude larger than those obtained from <sup>17</sup>O or <sup>13</sup>C NMR data [15]. The reason is that NMR, as opposed to viscosity, is directly sensitive to the internal motions of the molecules themselves. Therefore, the range of  $\tau_R$  values extracted from our data for the GdL<sub>2</sub> particles can be considered to be compatible with the one found for the adduct with HSA, the remaining difference being related to the larger size of the GdL<sub>2</sub> particles.

The NMRD-profile of a GdL<sub>2</sub> suspension is reported on Fig. 2 [11]. The high field peak in the range 20–60 MHz is a consequence of the slow rotational correlation time of the self-aggregates; indeed, superparamagnetic properties can be excluded in the present edifices. The decrease of  $\eta$  (and therefore  $\tau_R$ ) versus temperature displayed in Fig. 1 explains, at least partly, why  $r_1$  values are lower at 75 than at 25 °C. Apart of this, the decrease of  $r_1$  observed between 25 and 5 °C, is the signature of a relaxivity limited by a

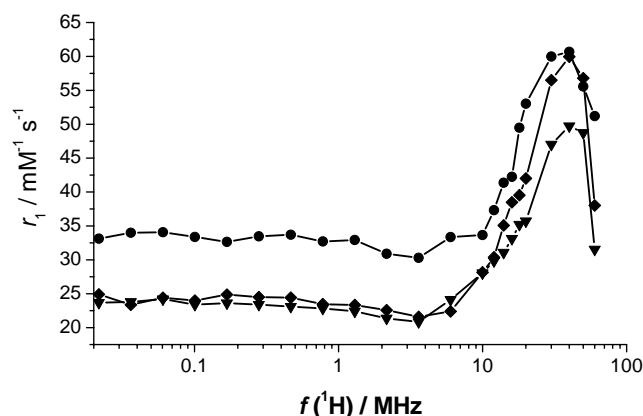


Fig. 2. NMRD profiles of GdL<sub>2</sub> at 5 °C (◆), 25 °C (●), and 75 °C (▼). [GdL<sub>2</sub>]<sub>tot</sub> =  $1.5 \times 10^{-3}$  M; pH 7.2 (pipes). Redrawn from [11].

slow water exchange rate [7,14]: the residence time is long compared to the relaxation time. Interest in large particles with contrast agent ability, such as those generated in GdL<sub>2</sub> suspensions, stems from their long retention time in blood, needed for angiographic applications. Another advantage of GdL<sub>2</sub> aggregates lies in the fact that the nanoparticles generate a large and almost constant relaxivity in the frequency range 30–50 MHz, which remains sizable at 63 MHz ( $30\text{--}50\text{ nm}^{-1}\text{ s}^{-1}$ ), which is important for biomedical applications. Many of the known supramolecular systems have a sharper relaxivity peak around 20 MHz followed by an abrupt drop at higher frequencies. Another potential asset of GdL<sub>2</sub> lies in the diameters of the two main kinds of nanoparticles found at concentrations feasible for imaging, 10 and 60 nm. This is well below 100 nm, a limit pertaining to intravenous injections since larger particles are easily retained by the liver. Finally, we have shown that addition of an equimolar quantity of Zn<sup>II</sup> does not affect the relaxivity [11].

In order to better assess the potentiality of GdL<sub>2</sub> nanoparticles for in vivo MRI analyses, we have determined the longitudinal ( $T_1$ ) and transverse ( $T_2$ ) relaxation times of the water protons of GdL<sub>2</sub> suspensions under MRI conditions. The MRI images (Fig. 3) were obtained under clinical experimental conditions by inserting a double-wheeled support fitted with 15 test tubes (1 cm diameter and 10 cm long) into one of the MRI scanners of the University Hospital. We have tested 10 different concentrations of GdL<sub>2</sub>, from  $10^{-5}$  M (tube 1) to  $10^{-3}$  M (tube 11) and compared these suspensions to water (tube 12), the Gd<sup>3+</sup> aquo-ion  $10^{-3}$  M (tube 13) and to two concentrations of the commercial contrast agent Omniscan<sup>®</sup> (tubes 14, 15). Qualitatively, the brightness of the spots generated by the GdL<sub>2</sub> suspensions increases with concentration to reach a maximum in the range  $(0.9\text{--}2.0) \times 10^{-4}$  M and then decreases with increasing concentration. The data confirm the laboratory results reported previously [11] in that a  $10^{-4}$  M suspension of GdL<sub>2</sub> has approximately the same effect than a  $5 \times 10^{-3}$  M solution of Omniscan<sup>®</sup>. More quantitatively, the relaxivity



20 MHz and 23 °C; in addition, they are relatively simpler to synthesize. Their good resistance to exchange with  $\text{Zn}^{\text{II}}$  and the absence of cytotoxicity [16] is also an asset. On the other hand, kinetic tests conducted in presence of large amounts of Arsenazo III or NaCl [17] have revealed a too great lability, related to the relatively low pLn value ( $\text{pEu} = 10$  [11] as compared to 15.4 for  $[\text{Eu}(\text{edta})]^-$  [18]) and this aspect must be improved before the  $\text{GdL}_2$  nanoparticles can be practically used.

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